

PREPARATION OF 3-(6-DEOXY- $\beta$ -D-GLUCOPYRANOSYLOXY) AND 3-(6-DEOXY- $\alpha$ -L-MANNOPYRANOSYLOXY)ANDROSTANE DERIVATIVES WITH UNSATURATED SIDE CHAIN IN POSITION 17 $\beta$ \*

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In our previous papers<sup>1-4</sup> we described the preparation of 3-( $\beta$ -D-glucopyranosyloxy) and 3-( $\beta$ -D-galactopyranosyloxy) derivatives of steroids with an  $\alpha,\beta$ -unsaturated ester chain in position 17 $\beta$  of androstane skeleton. In connection with this project we have studied silver silicate promoted<sup>5</sup> glycosylation of some of above mentioned steroidal derivatives with 2,3,4-tri-*O*-acetyl-6-deoxy- $\alpha$ -D-glucopyranosyl bromide and 2,3,4-tri-*O*-acetyl-6-deoxy- $\alpha$ -L-mannopyranosyl bromide. Glycosylation with the former gave corresponding  $\beta$ -D-quinovosides triacetates *I* – *III*, *V*, *VI*, and *IX* (see Table I).  $\beta$ -Configuration on anomeric centers in these compounds follow from *J*(1,2) of sugar moieties in proton NMR spectra; values in the region 7.5 – 8.0 Hz (cf. Table II) agree with those found in spectra of analogous  $\beta$ -D-glucopyranosides tetraacetates<sup>1-3</sup>.

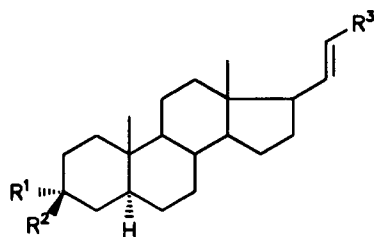
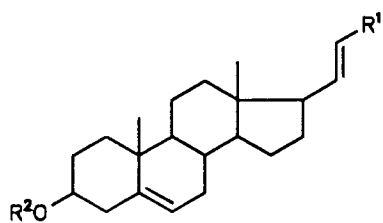
The glycosylation with the latter reagent gave  $\alpha$ -L-rhamnosides triacetates *IV*, *VII*, and *VIII*. In this case the value of *J*(1,2) of sugar moiety could not be used for the determination of configuration on anomeric center; according to the literature<sup>6</sup> *J*(1,2) of both  $\alpha$ - and  $\beta$ -methyl L-rhamnosides *XIX* and *XX* are identical. Thus, we used the substantial differences of chemical shifts of H-2, H-3, and H-5 (cf. Table III) and C-2, C-3, and C-5 (cf. Table IV). Both these criteria proved that our compounds have  $\alpha$ -configuration on anomeric center.

Hydrolysis of triacetates *I* – *IX* gave corresponding free  $\beta$ -D-quinovosides *X* – *XII*, *XIV*, *XV*, and *XVIII* and  $\alpha$ -L-rhamnosides *XIII*, *XVI*, and *XVII* (Table V).

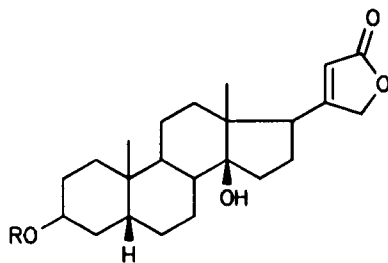
## EXPERIMENTAL

Melting points were determined on a micro melting point apparatus Boetius (Germany). Optical rotations were measured in chloroform at 25 °C. IR spectra (Tables VI and VII) were taken on a

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	R <sup>1</sup>	R <sup>2</sup>		R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>
<i>I</i>	COOCH <sub>3</sub>	Ac <sub>3</sub> -β-D-Qui	<i>V</i>	Ac <sub>3</sub> -β-D-Qui	H	CH <sub>3</sub>
<i>II</i>	COOC <sub>2</sub> H <sub>5</sub>	Ac <sub>3</sub> -β-D-Qui	<i>VI</i>	H	Ac <sub>3</sub> -β-D-Qui	CH <sub>3</sub>
<i>III</i>	C≡N	Ac <sub>3</sub> -β-D-Qui	<i>VII</i>	Ac <sub>3</sub> -α-L-Rha	H	C <sub>2</sub> H <sub>5</sub>
<i>IV</i>	COOC <sub>2</sub> H <sub>5</sub>	Ac <sub>3</sub> -α-L-Rha	<i>VIII</i>	H	Ac <sub>3</sub> -α-L-Rha	C <sub>2</sub> H <sub>5</sub>
<i>X</i>	COOCH <sub>3</sub>	β-D-Qui	<i>XIV</i>	β-D-Qui	H	CH <sub>3</sub>
<i>XI</i>	COOC <sub>2</sub> H <sub>5</sub>	β-D-Qui	<i>XV</i>	H	β-D-Qui	CH <sub>3</sub>
<i>XII</i>	C≡N	β-D-Qui	<i>XVI</i>	α-L-Rha	H	C <sub>2</sub> H <sub>5</sub>
<i>XIII</i>	COOC <sub>2</sub> H <sub>5</sub>	α-L-Rha	<i>XVII</i>	H	α-L-Rha	C <sub>2</sub> H <sub>5</sub>



*IX*, R = Ac<sub>3</sub>-β-D-Qui  
*XVIII*, R = β-D-Qui

β-D-Qui = 6-deoxy-β-D-glucopyranosyl

α-L-Rha = 6-deoxy-α-L-mannopyranosyl

Ac<sub>3</sub>-β-D-Qui = 2, 3, 4-tri-O-acetyl-6-deoxy-β-D-glucopyranosyl

Ac<sub>3</sub>-α-L-Rha = 2, 3, 4-tri-O-acetyl-6-deoxy-α-L-mannopyranosyl

Perkin-Elmer PE 580 spectrometer (wavenumbers in  $\text{cm}^{-1}$ ). Proton and carbon-13 NMR spectra were measured on Varian XL-200 instrument (FT mode, 200.06 and 50.31 MHz for  $^1\text{H}$  and  $^{13}\text{C}$ , respectively) at 23 °C in deuteriochloroform. For  $^1\text{H}$  NMR spectra tetramethylsilane was used as internal standard. Carbon-13 chemical shifts were referenced to the signal of solvent and recalculated to tetramethylsilane with relation  $\delta(\text{CDCl}_3) = 77.0$  ppm. The number of directly bonded hydrogen atoms was determined from the proton decoupled "attached proton test" spectra (APT, refs<sup>7,8</sup>). Chemical shifts are given in ppm ( $\delta$ -scale), coupling constants ( $J$ ) in Hz. All parameters were obtained by the first-order analysis. Column chromatography was performed on Silpearl (Kavalier, Votice, The Czech Republic) and thin-layer chromatography on silica gel G according to Stahl (ICN Biochemicals). Solutions in organic solvents were dried over anhydrous magnesium sulfate and the solvents were evaporated in vacuo (about 2 kPa). Analytical samples were dried over phosphorus pentoxide at 40 °C/26 Pa for 12 h. 2,3,4-Tri-*O*-acetyl-6-deoxy- $\beta$ -D-glucopyranosyl and 2,3,4-tri-*O*-acetyl-6-deoxy- $\alpha$ -L-mannopyranosyl bromides were prepared by passing dry hydrogen bromide into a solution of corresponding 6-deoxyhexose peracetate in dichloromethane. The preparations of starting hydroxy derivatives is described in refs<sup>3,9</sup>.

TABLE I

Yields and physico-chemical data of  $\beta$ -D-quinovosides triacetates *I* - *III*, *V*, *VI*, and *IX* and  $\alpha$ -L-rhamnosides triacetates *IV*, *VII*, and *VIII*

Compound	Yield, % solvent <sup>a</sup>	M.p., °C	$[\alpha]_D^{20}$ , (c) <sup>b</sup>	Formula M.w.	Calculated/Found	
					% C	% H
<i>I</i>	65	154 - 156	-10 (2.7)	C <sub>35</sub> H <sub>50</sub> O <sub>10</sub> 630.8	66.65	7.99
	E				66.45	8.23
<i>II</i>	51	154 - 155	-13 (2.7)	C <sub>36</sub> H <sub>52</sub> O <sub>10</sub> 644.8	67.06	8.13
	E				66.87	7.95
<i>III</i>	45	195 - 196	-11 (1.5)	C <sub>34</sub> H <sub>47</sub> NO <sub>8</sub> 597.8	68.32	7.93 <sup>c</sup>
	E + C				68.56	8.12
<i>IV</i>	32	154 - 156	-73 (1.1)	C <sub>35</sub> H <sub>52</sub> O <sub>10</sub> 644.8	67.06	8.13
	E + H				66.85	7.89
<i>V</i>	57	201 - 204	+6 (1.6)	C <sub>35</sub> H <sub>52</sub> O <sub>10</sub> 632.8	66.43	8.28
	E				66.68	7.97
<i>VI</i>	60	159 - 162	+5 (1.6)	C <sub>35</sub> H <sub>52</sub> O <sub>10</sub> 632.8	66.43	8.28
	H + E				66.72	8.56
<i>VII</i>	52	163 - 165	-4 (1.9)	C <sub>35</sub> H <sub>54</sub> O <sub>10</sub> 646.8	66.85	8.41
	E + M				67.09	8.67
<i>VIII</i>	51	140 - 142	-30 (1.5)	C <sub>35</sub> H <sub>54</sub> O <sub>10</sub> 646.8	66.85	8.41
	E + H				66.54	8.24
<i>IX</i>	30	248 - 250	0 (1.7)	C <sub>35</sub> H <sub>50</sub> O <sub>11</sub> 646.8	65.00	7.79
	E				65.08	8.05

<sup>a</sup> Solvents: C chloroform, E ether, H hexane, M methanol. <sup>b</sup> Measured in chloroform. <sup>c</sup> Calculated: 2.34% N; found: 2.45% N.

TABLE II  
 $^1\text{H}$  NMR spectral parameters (in  $\text{CDCl}_3$ ) of quinovoside triacetates I – III, V, VI, and IX. For other conditions see Experimental

Parameter	I <sup>a</sup>	II <sup>b</sup>	III	V <sup>a</sup>	VI <sup>a</sup>	IX <sup>c</sup>
Steroid unit						
H-18 (s, 3 H)	0.66	0.66	0.66	0.63	0.63	0.88
H-19 (s, 3 H)	0.99	0.99	0.99	0.77	0.78	0.96
H-21 (dd, 1 H)	5.79	5.77	5.28	5.78	5.78	4.88 <sup>d</sup>
H-20 (dd, 1 H)	6.96	6.94	6.70	6.94	6.95	–
H-3 (m, 1 H)	3.48 <sup>e</sup>	3.49 <sup>e</sup>	3.48 <sup>e</sup>	3.90 <sup>f</sup>	3.55 <sup>g</sup>	4.02 <sup>f</sup>
H-6 (1 H)	5.36 <sup>h</sup>	5.36 <sup>h</sup>	5.36 <sup>h</sup>	<i>i</i>	<i>i</i>	<i>i</i>
J(20,21)	15.6	16.0	16.5	16.0	16.0	–
J(17,20)	8.1	8.0	8.0	8.0	8.0	–
J(17,21)	1.2	1.2	1.0	1.2	1.0	≈0
Sugar unit						
H-1 (d, 1 H)	4.56	4.54	4.64	4.51	4.56	4.51
H-2 (dd, 1 H)	4.93	4.92	4.92	4.96	4.92	4.94
H-3 (t, 1 H)	5.16	5.17	5.16	5.17	5.15	5.17
H-4 (t, 1 H)	4.81	4.79	4.81	4.82	4.80	4.82
H-5 (dq, 1 H)	3.54	3.53	3.54	3.54	3.55 <sup>g</sup>	3.52
H-6 (d, 3 H)	1.23	1.23	1.23	1.23	1.23	1.23
J(1,2)	8.0	8.0	8.0	7.5	8.0	8.0
J(2,3)	9.5	9.5	9.5	9.5	9.5	9.5
J(3,4)	9.5	9.5	9.5	9.5	9.5	9.5
J(4,5)	9.5	9.5	9.0	9.5	9.5	9.5
J(5,6)	6.2	6.5	6.0	6.0	6.2	6.5
OAc (s, 3 H)	2.00	1.99	2.00	2.00	2.00	2.00
OAc (s, 3 H)	2.03	2.03	2.03	2.03	2.03	2.03
OAc (s, 3 H)	2.04	2.04	2.03	2.04	2.03	2.03

<sup>a</sup> Other signal: 3.72 s, 3 H ( $\text{COOCH}_3$ ). <sup>b</sup> Other signals: 4.18 q, 2 H and 1.29 t, 3 H ( $\text{COOCH}_2\text{CH}_3$ ,  $J = 7.2$ ). <sup>c</sup> Other signals: 2.75 m, 1 H (H-17); 5.87 X part of ABX system (H-22,  $J(\Lambda, X) \approx J(B, X) \approx 2$ ). <sup>d</sup>  $\Delta B$  part of  $\Delta BX$  system,  $J(\Lambda, B) = 18$ . <sup>e</sup>  $W = 32$ . <sup>f</sup>  $W = 14$ . <sup>g</sup> Multiplet of overlapped signals. <sup>h</sup> bd,  $J \approx 4.5$ . <sup>i</sup> Undeterminable value.

TABLE III  
 $^1\text{H}$  NMR spectral parameters (in  $\text{CDCl}_3$ ) of rhamnoside triacetates *IV*, *VII*, and *VIII*. For other conditions see Experimental

Parameter	<i>IV</i>	<i>VII</i>	<i>VIII</i>	<i>XIX</i> <sup>a</sup>	<i>XX</i> <sup>a</sup>
Steroid unit					
H-18 (s, 3 H)	0.66	0.63	0.63		
H-19 (s, 3 H)	1.01	0.78	0.81		
H-21 (dd, 1 H)	5.78	5.78	5.77		
H-20 (dd, 1 H)	6.94	6.94	6.94		
H-3 (m, 1 H)	3.49 <sup>b</sup>	3.87 <sup>c</sup>	3.53 <sup>b</sup>		
H-6 (1 H)	5.34 <sup>d</sup>	<sup>e</sup>	<sup>e</sup>		
<i>J</i> (20,21)	15.6	15.6	15.6		
<i>J</i> (17,20)	7.9	7.9	7.9		
<i>J</i> (17,21)	1.1	1.1	1.1		
$\text{COOCH}_2\text{CH}_3$ (t, 3 H)	1.29 <sup>f</sup>	1.29 <sup>g</sup>	1.29 <sup>g</sup>		
$\text{COOCH}_2\text{CH}_3$ (q, 2 H)	4.18 <sup>f</sup>	4.18 <sup>g</sup>	4.18 <sup>g</sup>		
Sugar unit					
H-1 (d, 1 H)	4.88	4.81	4.87	4.63	4.51
H-2 (dd, 1 H)	5.18	5.20	5.16	5.22	5.46
H-3 (dd, 1 H)	5.33	5.35	5.32	5.40	5.10
H-4 (bt, 1 H)	5.05	5.07	5.04	5.03	5.00
H-5 (dq, 1 H)	3.95	3.92	3.95	3.86	3.50
H-6 (d, 3 H)	1.20	1.20	1.20	1.21	1.30
<i>J</i> (1,2)	1.8	1.8	1.8	1.0	1.0
<i>J</i> (2,3)	3.4	3.5	3.4		
<i>J</i> (3,4)	10.1	10.1	10.2		
<i>J</i> (4,5)	9.9	9.7	9.7		
<i>J</i> (5,6)	6.2	6.2	6.3		
OAc (s, 3 H)	1.99	2.00	1.98		
OAc (s, 3 H)	2.04	2.06	2.04		
OAc (s, 3 H)	2.15	2.15	2.14		

<sup>a</sup> *XIX*, Methyl 2,3,4-tri-*O*-acetyl-6-deoxy- $\alpha$ -*D*-mannopyranoside; *XX*, methyl 2,3,4-tri-*O*-acetyl-6-deoxy- $\beta$ -*D*-mannopyranoside; values taken from literature<sup>6</sup>. <sup>b</sup>  $W \approx 32$ . <sup>c</sup>  $W \approx 10$ . <sup>d</sup> bd,  $J \approx 4.5$ . <sup>e</sup> Undeterminable value. <sup>f</sup>  $J = 7.1$ . <sup>g</sup>  $J = 7.2$ .

TABLE IV  
 $^{13}\text{C}$  NMR spectral parameters (in  $\text{CDCl}_3$ ) of  $\alpha$ -L-rhamnoside triacetates IV, VII, and VIII. For other conditions see Experimental

Carbon	IV	VII	VIII	XIX <sup>a</sup>	XX <sup>a</sup>
Steroid unit					
1	37.24	31.91	33.91		
2	29.25	24.69	28.61		
3	74.45	72.34	76.89		
4	38.25	34.13	36.98		
5	140.12	39.72	44.64		
6	121.28	28.42	29.04		
7	31.61	32.45	32.06		
8	31.61	35.49	35.47		
9	50.20	54.12	54.43		
10	36.71	35.90	35.64		
11	20.53	20.25	20.70		
12	37.19	37.40	37.48		
13	44.56	44.85	44.84		
14	55.99	55.75	55.78		
15	24.92	24.84	24.86		
16	26.96	27.00	26.99		
17	53.77	53.90	53.90		
18	12.96	13.22	13.20		
19	19.28	11.35	12.23		
20	150.41	150.75	150.62		
21	121.74	121.17	121.20		
C=O	166.54	166.66	166.60		
OCH <sub>2</sub> CH <sub>3</sub>	14.21	14.24	14.24		
OCH <sub>2</sub> C(CH <sub>3</sub> ) <sub>2</sub>	60.01	60.00	60.03		
Sugar unit					
1	95.59	95.37	95.39	98.1	98.7
2	71.27 <sup>b</sup>	71.37 <sup>b</sup>	71.34 <sup>b</sup>	69.3	86.6
3	69.08 <sup>b</sup>	69.29 <sup>b</sup>	69.13 <sup>b</sup>	69.0	70.7
4	70.39 <sup>b</sup>	70.58 <sup>b</sup>	70.58 <sup>b</sup>	70.4	70.5
5	66.20	66.35	66.17	66.0	69.6
6	17.30	17.32	17.34	17.3	17.3
OOCCH <sub>3</sub>	170.09	170.20	170.15	169.6	169.8
OOCCH <sub>3</sub>	169.89 <sup>c</sup>	170.05	169.94	169.6	169.4
OOCCH <sub>3</sub>	169.89 <sup>c</sup>	169.95	169.91	169.6	169.4
OOCCH <sub>3</sub>	20.87	20.93	20.91	20.5	20.6
OOCCH <sub>3</sub>	20.72	20.78	20.76	20.5	20.6
OOCCH <sub>3</sub>	20.66	20.73	20.70	20.5	20.4

<sup>a</sup> XIX, Methyl 2,3,4-tri-*O*-acetyl-6-deoxy- $\alpha$ -L-mannopyranoside; XX, methyl 2,3,4-tri-*O*-acetyl-6-deoxy- $\beta$ -L-mannopyranoside (values taken from literature<sup>10</sup>). <sup>b</sup> Signals are assigned tentatively and can be mutually interchanged. <sup>c</sup> Overlapped signals.

General Procedure for Preparation of Quinovosides Triacetates *I* – *III*, *V*, *VI*, and *IX*  
and Rhamnosides Triacetates *IV*, *VII*, and *VIII*

A dry mixture of a hydroxy derivative (0.5 mmol), silver silicate<sup>5</sup> (0.7 g) and ground molecular sieve 4 Å (1 g) was stirred in vacuo (10 Pa) for 4 h. The flask was then filled with argon under slight overpressure (about 5 kPa) and 1,2-dichloroethane (10 ml) was injected through a septum. The mixture was stirred at room temperature for 10 min and a solution of corresponding halogenose (494 mg, 1.4 mmol) in 1,2-dichloroethane (3 ml) was added. After stirring at room temperature for 20 h, the catalyst was removed by filtration through a column of silica gel layered with Celite. The column was washed with dichloromethane–ether (4 : 1) and the combined filtrates were evaporated in vacuo. The residue was chromatographed on a column of Silpearl (30 g) in mixture toluene–ether (9 : 1). Obtained product was further purified by crystallization. For yield, solvent used for crystallization, melting points, optical rotations, and elemental analyses see Table I.

TABLE V

Yields and physico-chemical data of  $\beta$ -D-quinovosides *X* – *XII*, *XIV*, *XV*, and *XVIII* and  $\alpha$ -L-rhamnosides *XIII*, *XVI*, and *XVII*

Compound	Yield, % solvent <sup>a</sup>	M.p., °C	[ $\alpha$ ] <sub>D</sub> , ° (c) <sup>b</sup>	Formula M.w.	Calculated/Found	
					% C	% H
<i>X</i>	88	205 – 207	–32	C <sub>29</sub> H <sub>44</sub> O <sub>7</sub>	69.02	8.79
	E		(0.8)	504.7	68.88	9.02
<i>XI</i>	87	188 – 192	–40	C <sub>30</sub> H <sub>46</sub> O <sub>7</sub>	69.47	8.94
	M + W		(0.7)	518.7	69.23	9.09
<i>XII</i>	85	254 – 259	–34	C <sub>28</sub> H <sub>41</sub> NO <sub>5</sub>	71.31	8.76 <sup>c</sup>
	C + M		(0.6)	471.6	71.05	8.53
<i>XIII</i>	84	207 – 211	–72	C <sub>30</sub> H <sub>46</sub> O <sub>7</sub>	69.47	8.94
	E + A		(1.1)	518.7	69.35	9.06
<i>XIV</i>	77	120	+16	C <sub>29</sub> H <sub>46</sub> O <sub>7</sub>	68.75	9.15
	H		(1.6)	506.7	68.46	8.92
<i>XV</i>	86	205 – 207	+24	C <sub>29</sub> H <sub>46</sub> O <sub>7</sub>	68.75	9.15
	E + H		(1.7)	506.7	68.56	8.87
<i>XVI</i>	88	212 – 215	–12	C <sub>30</sub> H <sub>48</sub> O <sub>7</sub>	69.20	9.29
	E + A		(1.0)	520.7	69.16	9.01
<i>XVII</i>	86	198 – 200	–26	C <sub>30</sub> H <sub>48</sub> O <sub>7</sub>	69.20	9.29
	A		(1.1)	520.7	69.89	8.97
<i>XVIII</i>	37	298 – 299	–7	C <sub>29</sub> H <sub>44</sub> O <sub>8</sub>	66.90	8.52
	E		(2.2)	520.7	67.05	8.75

<sup>a</sup> Solvents: A acetone, C chloroform, E ether, H hexane, M methanol, W water. <sup>b</sup> Measured in chloroform. <sup>c</sup> Calculated: 2.97% N; found: 2.68% N.

TABLE VI

IR spectra ( $\tilde{\nu}$ ,  $\text{cm}^{-1}$ ; in  $\text{CCl}_4$ ) of  $\beta$ -D-quinovosides triacetates *I* – *III*, *V*, *VI*, and *IX* and  $\alpha$ -L-rhamnosides triacetates *IV*, *VII*, and *VIII*. For other conditions see Experimental

Compound	C=O <sup>a</sup>	C=O <sup>b</sup>	C=C <sup>c</sup>	C–O
<i>I</i>	1 752	1 713	1 650	1 257, 1 080, 1 058, 1 038
<i>II</i>	1 760	1 719	1 652	1 247, 1 218, 1 035
<i>III</i> <sup>d</sup>	1 756	–	1 628	1 254, 1 078, 1 037
<i>IV</i> <sup>e</sup>	1 750	1 719	1 650	1 224
<i>V</i>	1 753	1 718	1 652	1 254, 1 078, 1 037
<i>VI</i>	1 754	1 714	1 650	1 252, 1 077, 1 036
<i>VII</i> <sup>e</sup>	1 753	1 719	1 652	1 225
<i>VIII</i> <sup>e</sup>	1 752	1 719	1 651	1 225
<i>IX</i> <sup>f</sup>	1 748	1 748	1 621	1 252, 1 232, 1 075, 1 035

<sup>a</sup> Carbonyl of glycoside acetates. <sup>b</sup> Carbonyl of unsaturated ester. <sup>c</sup> Conjugated double bond. <sup>d</sup> Other band 2 228 (C≡N). <sup>e</sup> Measured in chloroform solution. <sup>f</sup> Other bands 3 604, 3 408 (OH).

TABLE VII

IR spectra ( $\tilde{\nu}$ ,  $\text{cm}^{-1}$ ; KBr pellet) of  $\beta$ -D-quinovosides *X* – *XII*, *XIV*, *XV*, and *XVIII* and  $\alpha$ -L-rhamnosides *XIII*, *XVI*, and *XVII*. For other conditions see Experimental

Compound	O–H	C=O	C=C <sup>a</sup>	C–O
<i>X</i>	3 440	1 723, 1 705	1 652	1 072
<i>XI</i>	3 439	1 725	1 658	1 055, 1 018
<i>XII</i> <sup>b</sup>	3 403	–	1 628	1 066, 1 012
<i>XIII</i>	3 405	1 733	1 655	1 061, 1 031
<i>XIV</i>	3 445	1 723	1 650	1 170, 1 070
<i>XV</i>	3 433	1 727	1 653	1 066, 1 012
<i>XVI</i>	3 405	1 733	1 655	1 055, 1 034
<i>XVII</i>	3 420	1 725	1 653	1 067, 1 023
<i>XVIII</i>	3 456	1 757, 1 742	1 621	1 093, 1 023

<sup>a</sup> Conjugated double bond. <sup>b</sup> Other band 2 222 (C≡N).



General Procedure for Preparation of Quinovosides *X – XII*, *XIV*, *XV*, and *XVIII*  
and Rhamnosides *XIII*, *XVI*, and *XVII*

To a solution of a glycoside triacetate (0.1 mmol) in methanol (5 ml) a solution of sodium methoxide in methanol (5%, 5 drops) was added. After stirring for 2 h at room temperature solid carbon dioxide (about 100 mg) was added and the solvent was evaporated in vacuo. The residue was chromatographed on a column of Silpearl (30 g) in chloroform–methanol (9 : 1). Obtained product was purified by crystallization. For yield, solvent used for crystallization, melting points, optical rotations, and elemental analyses see Table V.

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